

1 **ISCEV extended protocol for the dark-adapted full-field ERG b-**
2 **wave transfer (intensity-response) function**

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19 **Abstract**

20
21 The International Society for Clinical Electrophysiology of Vision (ISCEV) standard for full-field
22 electroretinography (ERG) describes a minimum procedure but encourages more extensive testing.
23 This ISCEV extended protocol describes an extension of the ISCEV standard, in which methods to
24 record and evaluate the growth of the dark-adapted (DA) ERG b-wave with increasing stimulus
25 strength are described. The protocol includes use of the lowest flash strength required to generate a
26 reliable DA ERG b-wave and a series of flashes of increasing strengths used to define the maximal b-
27 wave amplitude. The ERG transfer (“intensity-response”) function can more comprehensively
28 characterize generalized rod system function than the ISCEV-standard ERG protocol, and may be of
29 diagnostic or prognostic value in disorders that cause generalized rod system dysfunction.

30
31 Keywords. Clinical standards; Electroretinogram (ERG); Full-field ERG; International Society for
32 Clinical Electrophysiology of Vision (ISCEV); Dark-adapted (DA); Intensity-response (IR);
33 Retinopathy; Naka-Rushton

38 **Introduction**

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40 The International Society for Clinical Electrophysiology of Vision (ISCEV) standard for full-field
41 electroretinography (ERG) describes a minimum set of tests but encourages the use of additional
42 ERG protocols for clinical ERG testing.(1) This extended protocol describes the dark-adapted (DA)
43 ERG transfer function, historically referred to as the “intensity-response” (I-R) function, a
44 specialized procedure that is well established and broadly accepted by experts in the field. This
45 protocol was prepared by the authors in accordance with ISCEV procedures
46 (<http://www.iscev.org/standards/index.html#guide2procedures>).

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48 **Scope and Applications**

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50 The ERG is a graded response, i.e., its amplitude, timing and waveform change with increasing
51 stimulus strength (Fig 1). This protocol describes the process of recording dark-adapted ERGs
52 using a series of increasing stimulus strengths, and of analyzing the data by their fit to a heuristic
53 model. The derived parameters of the model characterize the maximal rod-mediated retinal
54 response and provide a measure of retinal sensitivity that may aid understanding of
55 pathophysiology in some retinopathies.

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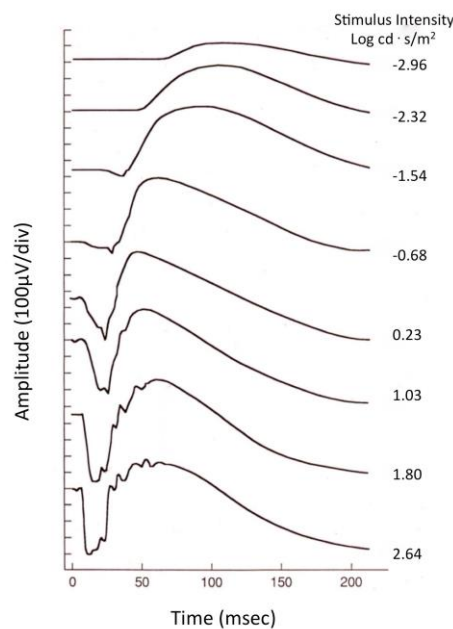
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69 *Figure 1. Human ERG waveforms measured over a > 5 log unit range of stimulus strengths. Figure*
70 *initially appeared in Principles and Practice of Clinical Electrophysiology of Vision. Reprinted with*
71 *permission of Mosby Year Book, Inc.(3)*

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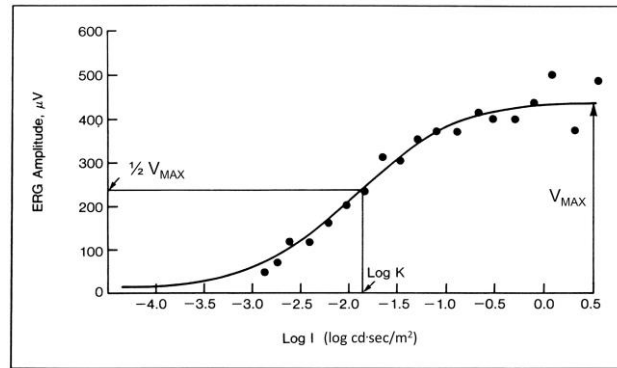


Figure 2. DA ERG b-wave as a function of stimulus strength.

Naka and Rushton(4) were the first to model the growth of the ERG b-wave with increasing stimulus strength. Figure 2 is an example of b-wave amplitude data recorded as a function of log flash strength in a normal human observer.

They showed that the hyperbolic function

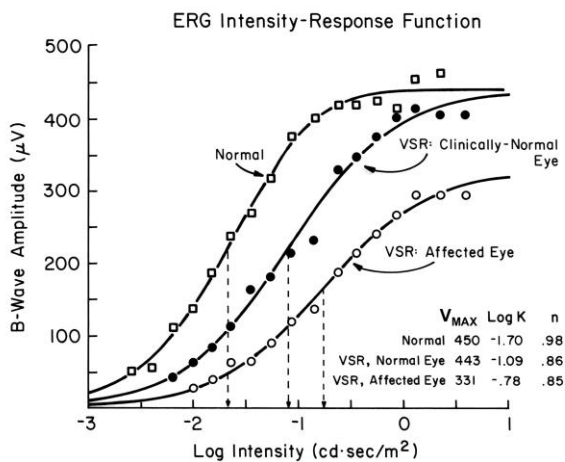
$$V = V_{\max} I^n / I^n + K^n$$

where V (μV) is the ERG b-wave amplitude generated in response to flash strength I ($\text{cd}\cdot\text{s}/\text{m}^2$), provided a reasonable fit to b-wave amplitudes throughout most of the stimulus range. The derived parameter V_{\max} (μV) is the asymptotic amplitude of the function, K ($\text{cd}\cdot\text{s}/\text{m}^2$) is the flash strength that elicits a response that is $1/2 V_{\max}$, and n is a dimensionless number representing the slope of the curve and generally considered to equal 1. V_{\max} has been interpreted as an index of both the number of rods responding and the gain ($\mu\text{V}/\text{quanta}$) for each b-wave generator. A reduction in V_{\max} can occur secondary to loss of photoreceptors, disruption of the dark current, inner retina dysfunction or some other type of response compression. The parameter K has been interpreted as an index of retinal sensitivity that represents the efficiency of quantal capture. An elevation in K would shift the entire IR function to the right, indicating that a stronger stimulus is required to elicit b-waves of comparable amplitude. Reductions in V_{\max} and elevations in K may be seen individually, or more frequently, in combination in many retinal diseases. Figure 2 is an example of b-wave amplitude data recorded as a function of log flash strength in a normal human observer.

105 Intensity-response function parameters V_{max} and K may be used to obtain additional information
 106 about the etiology or prognosis in a number of disorders. Naka-Rushton parameters have helped
 107 characterize fundamental differences between the mechanisms of rod dysfunction and
 108 degeneration in rod-cone dystrophy (retinitis pigmentosa; RP) and cone-rod dystrophy (18).
 109 Patients with RP typically show a loss in V_{max} along with an elevation in K , whereas patients with
 110 cone-rod dystrophy usually show normal values for these parameters. An important exception to
 111 this pattern is KCNV2-retinopathy (“cone dystrophy with supernormal rod ERG”), characterized by
 112 generalized cone dysfunction and pathognomonic DA ERG changes and an abnormal ERG I-R
 113 function. (45, 51)

114 Figure 3 shows an illustrative example of a normal I-R function compared with a case of central
 115 retinal vein occlusion (CRVO). There is loss in V_{max} in the affected eye of 0.13 log but an elevation in
 116 K of 0.92 log. The fellow eye also showed a large elevation in K (0.61 log) with a normal V_{max} ,
 117 highlighting the possibility of subclinical involvement.

118



129
 130 *Figure 3. ERG intensity-response functions in the affected and clinically normal eyes of a patient with*
 131 *venous-stasis retinopathy, compared to an age-similar normal subject.*

132
 133 Intensity-response functions also have been used to evaluate the timeline of retinal development
 134 and aging (33,47,56,62), and toxicity and efficacy in pharmaceutical studies (35,39,48,49,50). They
 135 have been recorded in many degenerative retinal disorders (10,14,16-20,26,34,55,58), as well as in
 136 congenital stationary night blindness (CSNB), in which they revealed differences between the
 137 complete and incomplete forms (64). The IR function may also be of use in other disorders such as

138 age-related macular degeneration (AMD; 12), altitude retinopathy (44), central retinal artery and
139 vein occlusions (8,9,21,27,31,32) and diabetic retinopathy (7,59,63).

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141 Patients who have an elevation in log K will also have a delay in b-wave timing, because peak times
142 change with stimulus strength, and thus, retinal sensitivity to that stimulus level. ERG peak time
143 measurements can be used to estimate retinal sensitivity loss, and have been used to predict
144 proliferative retinopathy in CRVO (8, 31, 32) and diabetic retinopathy.(7)

145

146 **Patient Population**

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148 Patients of all ages able to tolerate ganzfeld stimulation, referred for investigation of rod-mediated
149 retinal function. Using this paradigm, patients with cone-mediated abnormalities will usually show
150 minimal changes in the derived parameters produced by the curve fit to the data. (14)

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153 **Technical Issues**

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155 This protocol has the same requirements as those outlined under the Basic Technology section of
156 the ISCEV ERG protocol [1]. Additional considerations are outlined below.

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158 a) Range of flash strengths. To adequately characterize the I-R function, flash stimuli must span a
159 range that includes the lowest flash strength required to generate a reliable DA ERG b-wave and
160 that required to generate a maximal b-wave amplitude. This normally occurs over a range of 3.5 to
161 4 log units of flash strengths.

162

163 b) Inter-stimulus interval. The inter-stimulus interval should be sufficiently long to maintain the
164 same level of dark adaptation throughout the procedure. The ISCEV ERG standard specifies an
165 inter-stimulus interval of 2 seconds for DA 0.01 and 10 seconds for DA 3.0, but there are no specific
166 recommendations for stimuli between these 2 stimulus strengths. It is recognized that the IR
167 function will asymptote at stimuli lower than DA 3.0 for most individuals.

168

169 c) Amplifier gain. Amplifier gain will need to be higher for the dim stimuli, and should be increased
170 until responses can be seen well enough to judge reliability.

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172 d) Signal averaging. Averaging is not usually required for generating waveforms for IR analysis as
173 long as responses are replicable. Small waveforms or responses of long peak time may be prone to
174 noise or intrusion of blink and eye movement artifacts.

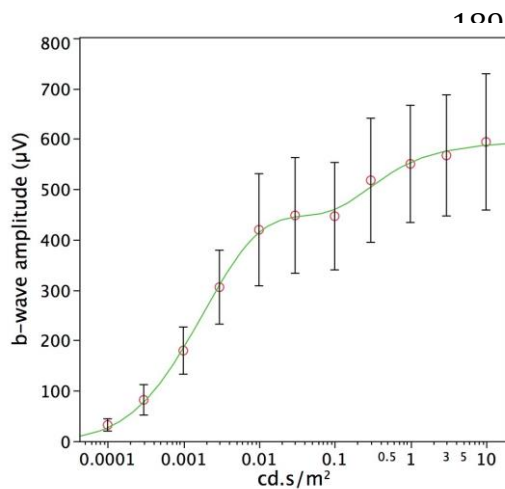
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176 e) Fitting the function. Near the asymptote of the IR function, a second hyperbolic function can be
177 seen in normal subjects (see fig. 4). By showing that this is observed in rod monochromats, Peachey
178 et al. (22) suggested that the second limb did not result from an interaction between rod and cone
179 systems but is more likely to represent destructive interference between the processes responsible
180 for the a- and b-waves, since b-wave amplitudes are measured from the trough of the a-wave to the
181 peak of the b-wave.

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183 f) Nomenclature of the function. In seminal studies and older publications the ERG “intensity-
184 response” function is used to describe the ERG stimulus-response series or ERG transfer function. It
185 is acknowledged that flashes should be described in terms of strength rather than intensity but the
186 widely used term “intensity-response” (I-R) function is retained in reference to historical data and
187 older publications.

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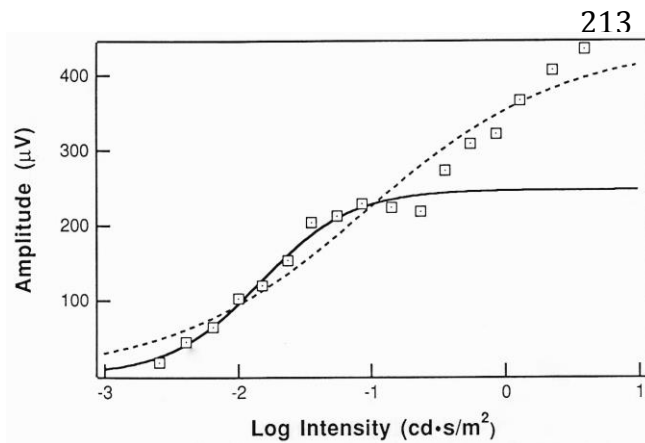
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200 *Figure 4. An IR function recorded from normal subjects (n=85): Circles are averages, error bars the*
201 *95% confidence limits and the green line a mathematical model that comprises the 2nd limb, seen*
202 *above flash strengths of approximately 0.1 cd.s./m².*

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The occurrence of a 2nd limb may confound standard application of the Naka-Rushton function, because the data variation does not follow a single hyperbolic function. If a single Naka-Rushton equation is forced to fit all of the data, the result will be a spuriously high V_{max} and an elevated $\log K$. The latter is illustrated in fig 5, which shows fits of the Naka-Rushton function to data obtained from one subject. When all of the data are fit together (dashed line), estimates of V_{max} increased from 233 μ V to 438 μ V, and the corresponding $\log K$ estimates increased from -1.89 to -1.03, when compared to the parameters obtained from fitting just the 1st limb (solid line).



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Figure 5. B-wave intensity-response data, fit by single Naka-Rushton equations to all of the data (dashed line) and to just the first limb (solid line). Figure reprinted with permission of Documenta Ophthalmologica.(5)

229 A heuristic method for identifying and excluding the 2nd limb has been described(5). The optimal
230 stimulus increment size for recognizing the 2nd limb was 0.4 log unit up to about the point the b-
231 wave begins to grow rapidly, and 0.2 log unit steps afterward. The smaller increment is necessary
232 to recognize the occurrence of a 2nd limb when it exists. The 2nd limb can also be excluded manually,
233 i.e. those data points that do not form a part of the single hyperbolic function can be omitted prior
234 to Naka-Rushton fit.

235

236 Calibration

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238 All stimuli should be individually calibrated and rechecked over time. Nominal flash strengths or
239 nominal increments in flash strengths should not be used. Standard ERG stimuli have a very short
240 duration (10 μ s for xenon bulbs), so a calibration device that can time-average flashes is required.
241 Low strength flashes from a xenon source generally vary more than flashes from LEDs and may
242 require assessment of multiple flashes to measure mean flash strength. LED sources can produce
243 more reliable flash stimuli because their output is determined by the current applied to them. Most
244 conventional ERG equipment that is manufactured currently use LEDs, which produce more stable
245 flash strengths.

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249 **Protocol Specifications**

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251 This protocol follows the same procedures for patient preparation and recording that are outlined
252 under the Clinical Protocol section of the ISCEV ERG protocol(1). Other specifications are listed
253 below.

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255 a) Flash strengths. Flash stimuli should span a range of approximately 3.5 to 4.0 log units. In the
256 absence of dysfunction the typical range would be approximately -3.5 to 0.5 log $\text{cd}\cdot\text{s}/\text{m}^2$, starting
257 with the lowest flash strength that will generate a reliable ERG b-wave ($>10 \mu\text{V}$) up to a flash
258 strength that generates a maximal b-wave response. Initially, stimuli should be recorded in
259 increasing steps of about 0.4 log units, until the b-wave amplitude begins to grow rapidly (near log
260 K). Thereafter, we recommend that step sizes be reduced to 0.2 – 0.3 log units, if the data are to be
261 fit by a single saturating function. Thus, at a minimum, about a dozen points should be recorded in a
262 normally-sighted subject.

263

264 b) Inter-stimulus interval. To avoid light adaptation this protocol specifies a time between flashes of
265 2 seconds up to $0.01 \text{ cd}\cdot\text{s}/\text{m}^2$; at least 3 seconds for stimuli up to $0.1 \text{ cd}\cdot\text{s}/\text{m}^2$; 5 seconds for stimuli
266 up to $3 \text{ cd}\cdot\text{s}/\text{m}^2$, and 10 seconds for higher flash strengths.

267

268 c) Amplifier gain. There is no specific requirement for amplifier gain except that it needs to be high
269 enough to evaluate the waveform and may need to be increased for responses to dim stimuli.

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271 d) Signal averaging. If averaging is needed or desired, a small number of 3 to 10 sweeps are usually
272 sufficient. Care must be taken to exclude spurious signals, such as eye movement and blink
273 artefacts, from averaged responses.

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276 **Response Evaluation**

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278 The DA ERG b-wave amplitudes should be measured as described in the ISCEV ERG standard (1).
279 The b-wave amplitudes at different flash intensities should then be input into one of the many
280 commercially or privately available computer programs that provide a fit to the data using a Naka-
281 Rushton-type function. Many of these programs use the Michaelis-Menten equation for enzyme
282 kinetics. The Michaelis-Menten equation is the same as the Naka-Rushton equation except that it
283 assumes a slope (n) of 1. The program used, in addition to fitting a curve to the data, should also
284 provide estimates of the maximum amplitude (V_{max}) and the semi-saturation constant (K). The plot
285 of b-wave amplitude vs. flash strength should be visually examined to determine if there is a second
286 limb, and the computer fit to the data should be adjusted either by omitting the points on the limb
287 and refitting the data, or by using a heuristic method that fits both limbs and eliminates the second
288 (5).

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290 **Reporting**

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292 In some cases a qualitative description of the I-R function may be sufficient to corroborate or
293 suggest a diagnosis e.g. abnormal I-R in KCNV2-retinopathy. Parameters V_{max} and log K should be
294 reported, as well as normal ranges for fully dilated eyes. It is recognized that for some applications,
295 a qualitative description of the I-R function may be sufficient to corroborate or suggest a diagnosis.
296 Eyes with smaller pupils will have an increasingly dimmer retinal illuminance (measured in
297 trolands), which will affect the value for log K. For this reason, pupil size should always be
298 measured. Compensation for light attenuation from small pupils is possible using table 1, and any
299 correction to log K should be clearly acknowledged.

300

If the pupil diameter is:	Then the pupil log area is:	And this number should be added to the value of log K:
8 mm	1.70	0
7 mm	1.59	-0.11
6 mm	1.45	-0.25
5 mm	1.29	-0.41
4 mm	1.10	-0.60

Table 1. The effect of pupil size on estimates of log K.

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303 10. Justification for the Protocol Details

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305 A literature review was performed using the Medline search engine, for human studies that used
 306 the words ERG and intensity response or Naka-Rushton or Michaelis-Menten or luminance
 307 response. Studies dealing with the photopic hill, photopic negative responses (PhNRs), wavelets
 308 and multifocal ERGs were excluded. Table 2 shows the results of this search.

309

310 Of 57 studies evaluated (Table 2), 60% used a flash strength range of 3.5 to 4.5 log units (range =
 311 2.5 - 7.5). Forty-seven percent used a flash step size of either 0.2 or 0.25 log, but there was a
 312 spread of data, shown in figure 6. The larger step sizes could easily misidentify a 2nd limb unless
 313 lower luminances were used. Most studies used the Naka-Rushton function to analyze the data.

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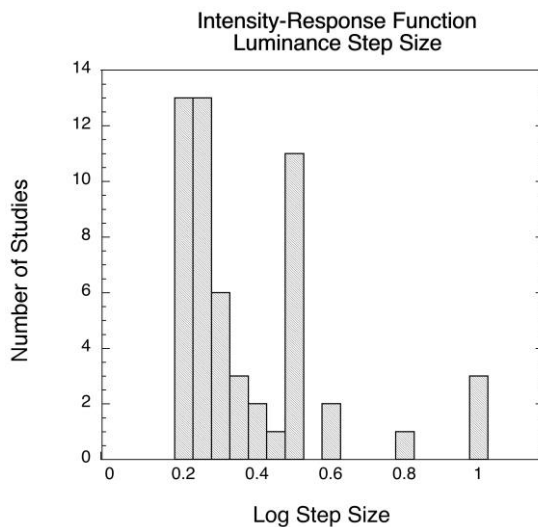


Figure 6. Luminance step sizes used for collection of ERG “intensity-response” data.

330 Table 2. Literature used for analysis.
331

Citation	Luminance Range	Luminance Step Size	Method of Fitting	Patient Population
(9)	6 log (-2.58 to 3.58 log cd·s/m ²)	0.5 log	none	Central Retinal Artery Occlusion
(10)	3 log	15 steps (0.2 log)	Naka-Rushton logistic function	RP 27 patients after renal transplant
(11)	5 log	11 steps (0.5 log)	Naka-Rushton	AMD
(12)	3.7 log (-2.9 to 0.6 log cd·s/m ²)	15 steps (0.25 log)	Naka-Rushton	RP
(13)	3.25 log	16 steps (0.25 log)	Naka-Rushton	Isotretinoin toxicity
(14)	4.0 log (-2 to 2 log scot td-s)	6 steps (0.6 log)	Naka-Rushton	Supernormal rod ERG
(15)	3.5 log	0.5 log or 1.0 log	none	RP
(16)	7.5 log	0.2 log per step	Naka-Rushton	RP, Cone Dystrophy
(17)	4.1 log (using rod isolation)	0.2 log per step	Naka-Rushton	RP
(18)	4.1 log (using rod isolation)	0.2 log per step	Naka-Rushton	RP
(19)	4.1 log (using rod isolation)	18 steps (0.2 log)	Naka-Rushton	X-linked RP
(20)	3.5 log (-2.61 to 0.87 log cd·s/m ²)	8 steps (0.5 log)	Naka-Rushton	CRVO
(21)	4 log	17 steps (0.25 log)	Naka-Rushton	Normal and Achromatopsia
(22)	4 log	18 steps (0.2 log)	Naka-Rushton	Albinism
(23)	3.8 log (-2.97 to 0.82 log cd·s/m ²)	13 steps (0.3 log)	Naka-Rushton	Cone-Rod Degeneration
(24)	4 log	18 steps (0.2 log)	Naka-Rushton	Sickle Cell Retinopathy
(25)	3.8 log (-2.97 to 0.82 log cd·s/m ²)			Elevated Cyclic Guanosine Monophosphate-Type Human Retinal Degeneration
(26)	3.5 log (-1.5 to 2 log scot td-s)	13 steps (0.25 log)	Naka-Rushton	Central Retinal Vein Occlusion
(27)	3.6 log	8 steps (0.6 log)	Naka-Rushton	Normal
(28)	4.5 log (-3.6 to 0.9 log cd·s/m ²)	10 steps (0.45 log)	Naka-Rushton	Normal
(29)	5 log (-1 to 4.0 log scot td-s)	19 steps (0.26 log)	Naka-Rushton	Normal
(30)	3.6 log	13 steps (0.27 log)	Naka-Rushton of 1st limb	Normal
(31)	4 log	0.2 log per step	Naka-Rushton	Central Retinal Vein Occlusion
(32)	Threshold to 0.29 log cd·s/m ²	0.2 log	Naka-Rushton	Central Retinal Vein Occlusion
(33)	2.5 log (-2.0 to 0.5 log scot td-s)	0.28 log	Naka-Rushton	Normal over the lifespan
(34)	3.25 log (-1.19 to 2.04 log scot td-s)	unspecified	Naka-Rushton	RP and Normal
(35)	3.50 log (-4.5 to -1 log cd·s/m ²)	0.2 log per step	Naka-Rushton	Sildenafil toxicity
(36)	3.25 log (-4.25 to -1 log cd·s/m ²)	14 steps (0.25 log)	Naka-Rushton	Normal
(37)	5.0 log (to 25.2 cd·s/m ²)	0.3 (Boston site); 1 log (Cambridge site)	Naka-Rushton	Normal Children and Adults

(38)	3.7 log (-0.7 to 3.0 log scot td-s) using rod isolation	unspecified	Naka-Rushton	Cone Dystrophy
(39)	3.50 log (-4.5 to -1 log cd·s/m ²)	0.2 log per step	Naka-Rushton	Sildenafil toxicity
(40)	4.0 log (-5.01 to -.96 log cd·s/m ²) blue light	11 steps (0.36 log)	Naka-Rushton	Normal - circadian rhythm
(41)	4.0 log (-3.95 to 0.05 log cd·s/m ²)	0.3 log; # of steps depending on start of 2nd limb	Naka-Rushton	Depression
(42)	3.25 log (-4.25 to -1.0 log cd·s/m ²) green light	14 steps (0.23 log)	Naka-Rushton	Normal-dilated vs undilated
(43)	3.25 log (-4.25 to -1.0 log cd·s/m ²) green light	14 steps (0.23 log)	none	Normal
(44)	4.5 log (-4.5 to 0.03 log cd·s/m ²)	11 steps (0.40 log)	Naka-Rushton	toxicity
(45)	4.06 log (-3.0 - 1.06 log cd·s/m ²)	8 steps (0.5 log)	none	KCNV2 Retinopathy
(46)	3.25 log (-4.25 to -1.0 log cd·s/m ²)	14 steps (0.23 log)	sigmoid curve	Normal, Patients with seasonal affective disorder
(47)	4.9 log (-3.27 to 2.16 log scot td-s) using rod isolation	0.4 log	Naka-Rushton	Preterm infants with and without ROP
(48)	3.5 log (-3.27 to 0.26 log cd·s/m ²)	8 steps (0.5 log)	Naka-Rushton	toxicity (bevacizumab)
(49)	4 log (-3.62 to 0.38 log cd·s/m ²)	4 steps (1.0 log)	Naka-Rushton	Patients (unknown diagnosis) receiving bevacizumab
(50)	4.9 log (-1.95 to 2.95 log scot td-s)	0.3 log	Naka-Rushton	Normal
(51)	5 log (-4 to 1 log cd·s/m ²)	10 steps (0.5 log)	Naka-Rushton	KCNV2 Retinopathy
(52)	5 log (-4 to 1 log cd·s/m ²)	10 steps (0.5 log)	Naka-Rushton	Normal
(53)	4.2 log (-0.7 to 3.5 log scot td-s)	7 steps (0.5 log)	H2 clearance curves	Normal
(54)	5.5 log (-4.5 to 1.0 log cd·s/m ²)	0.5/0.25 log	Naka-Rushton	Glaucoma/Normal (35/17) RP
(55)	3.5 log (-1.5 to 2 log scot td-s)	13 steps (0.27 log)	Naka-Rushton	Heterozygotes/Normal (11/19)
(56)	4 log (-3 to 1 log cd·s/m ²)	0.25 (adult)	Naka-Rushton	Infant ROP/Adult Normal (19/3)
(57)	3.9 log (-3.95 to -0.05 log cd·s/m ²)	0.5 (infant)	Naka-Rushton	Normal (19/3)
(58)	not specified	not specified	Naka-Rushton	Normal (10) RP/CRD/Normal (11/17/50)
(59)	3.1 log (-3.1 to 0 log cd·s/m ²)	variable (0.35 to 0.82 log)	Naka-Rushton	Diabetics/Normal (65/10)
(5)	threshold to 0.6 log cd·s/m ²	0.2 log	Naka-Rushton	CRVO/Normal (94/124)
(60)	3.8 log (-3.0 to 0.8 log cd·s/m ²)	18 steps (0.2 log)	Naka-Rushton	Normal (30)
(61)	3.0 log (-2.9 to 0.1 log cd·s/m ²)	0.3-0.4 log	Naka-Rushton	Normal (45: 61 eyes)
(62)	not specified	7 steps	Naka-Rushton	Normal (269)
(63)	4.0 log (-3.8 to 0.2 log cd·s/m ²)	0.2 log	1. Naka-Rushton 2. NR to first limb 3. Log model (-3.8 to -2.2)	Diabetics/Normal (152/40)

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